# Dose Adaptation of Antineoplastic Drugs in Patients with Liver Disease

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### **Abstract**

Dose adaptation for liver disease is important in patients treated with antineoplastic drugs because of the high prevalence of impaired liver function in this population and the dose-dependent, frequently serious adverse effects of these drugs. We classified the antineoplastic drugs marketed in Switzerland at the end of 2004 according to their bioavailability and/or hepatic extraction to predict their kinetic behaviour in patients with decreased liver function. This prediction was compared with kinetic studies carried out with these drugs in patients with liver disease. The studies were identified by a structured, computer-based literature search.

Of the 69 drugs identified, 52 had a predominant extrarenal (in most cases hepatic) metabolism and/or excretion. For 49 drugs, hepatic extraction could be calculated and/or bioavailability data were available, allowing classification according to hepatic extraction. For 18 drugs, kinetic studies have been reported in patients with impaired liver function, with the findings generally resulting in quantitative recommendations for adaptation of the dosage. In particular, recommendations are precise for 16 drugs excreted by the bile (e.g. doxorubicin and derivatives and vinca alkaloids). Validation studies comparing such recommendations with kinetics and/or dynamics of antineoplastic drugs in patients with decreased liver function have not been published.

We conclude that there are currently not enough data for safe use of cyctostatics in patients with liver disease. Pharmaceutical companies should be urged to provide kinetic data (especially hepatic extraction data) for the classification of such drugs and to conduct kinetic studies for drugs with primarily hepatic metabolism in patients with impaired liver function to allow quantitative advice to be given for dose adaptation.

Dose adaptation for patients with liver disease is more difficult to perform than for patients with impaired renal function. The main reason for this statement is that, unlike the creatinine clearance for the kidney, there is no *in vivo* surrogate to predict hepatic drug clearance. Due to the lack of such *in vivo* markers, predictions concerning dose adaptation in patients with liver disease can only be made based on the kinetic properties of the drugs to be administered and on kinetic studies of the drugs in patients with liver disease.<sup>[1]</sup>

Several reviews have covered this subject during the last few years.<sup>[1-5]</sup> In these reviews, drugs are listed according to hepatic extraction (E), which is an important determinant of the hepatic clearance of drugs. The hepatic clearance (Cl<sub>hep</sub>) of a drug can be expressed as (equation 1):

expressed as (equation 1): 
$$Cl_{hep} = \frac{(f_u \times Cl_i) \times Q}{(f_u \times Cl_i) + Q}$$
 (Eq. 1)

where Q is the blood flow across the liver, f<sub>u</sub> is the unbound fraction (free) of the drug and Cl<sub>i</sub> is the intrinsic clearance of the drug. Cl<sub>i</sub> represents the maximal capacity of the liver to metabolise a given drug, not taking into account limitations by liver perfusion. <sup>[6]</sup> Cl<sub>i</sub> can therefore reach values which are

The basis of the classifications used can best be understood by considering the extremes of equation 1, namely  $(f_u \times Cl_i) >> Q$  or  $Q >> (f_u \times Cl_i)$ . When  $(f_u \times Cl_i) >> Q$ , the denominator in equation 1 simplifies to  $(f_u \times Cl_i)$ , and  $Cl_{hep}$  equals (equation 2):

larger than Q.

$$Cl_{hep} = Q$$

(Eq. 2)

For these drugs, the liver has a very large metabolic capacity, E is approaching 1 and the blood flow across the liver becomes rate-limiting for hepatic clearance. The drugs are therefore called 'flow-limited', 'high capacity', 'high clearance' or 'high extraction'. Because of their high hepatic extraction, these drugs have a low bioavailability.

Since portal blood flow can be decreased in patients with liver cirrhosis or patients with multiple metastases, <sup>[7,8]</sup> hepatic clearance of such drugs is decreased in these situations, possibly necessitating a reduction of the maintenance dose. A second po-

tential problem of these drugs is an increase in their bioavailability in patients with porto-systemic shunts. Porto-systemic shunts are usually present in patients with portal hypertension due to liver cirrhosis or fibrosis or, of importance in patients with cancer, in patients with multiple metastases. [9,10] Therefore, when these drugs are administered orally in patients with portal hypertension, the initial and the maintenance doses have to be reduced according to the expected increase in bioavailability and to the decrease in hepatic blood flow. For intravenous administration, only the maintenance dose has to be reduced according to hepatic blood flow. A list of such drugs is given in a recent publication. [1]

For the second type of drugs,  $Q \gg (f_u \times Cl_i)$ , the metabolic capacity of the liver is much lower than blood flow across the liver. Equation 1 therefore simplifies to (equation 3):

$$\hat{Cl}_{hep} = (f_u \times Cl_i)$$

(Eq. 3)

These drugs are therefore called 'low extraction', 'low clearance' or 'capacity-limited'. They have a low extraction during the first passage across the liver and therefore have a high bioavailability, if bioavailability is not limited by other processes than first pass hepatic metabolism. Since Cl<sub>i</sub> decreases for most drugs in patients with liver cirrhosis due to a decrease in the activity of cytochrome P450 (CYP) isozymes<sup>[11,12]</sup> and/or glucuronyl transferases,<sup>[13-15]</sup> the maintenance dose of such drugs should generally be decreased in these patients. For drugs with a high binding to albumin (>90%), the situation may be more complex. The fu and the free concentration of these drugs can increase in patients with a low serum albumin level, e.g. patients with liver cirrhosis or malnourished patients, such as patients with cancer. An increase in the free concentration and/or fu of these drugs may be associated with increased toxicity and also, as shown in equation 3, with an increased hepatic clearance. [16,17] The actual hepatic clearance of these drugs is therefore difficult to predict in patients with chronic liver disease.

In between these two extremes, there are drugs with an 'intermediate extraction', showing characteristics of both groups. The dosage advice for these drugs in patients with liver cirrhosis is to start with a low dose and to up-titrate carefully to find the correct maintenance dose.

Table I. Classification of liver disease and severity of liver dysfunction

Parameter	Pathophysiological condition and clinical significance	Severity <sup>a</sup>
Alanine aminotransferase level	Breakdown (necrosis or apoptosis) of hepatocytes. Hepatocellular injury <sup>b</sup> if >2 × ULN	Moderate injury: 2-5 × ULN Severe injury: >5 × ULN
Alkaline phosphatase level	Cholestasis <sup>c</sup> if >2 × ULN	Moderate cholestasis: 2–5 × ULN Severe cholestasis: >5 × ULN
Serum bilirubin level	Cholestasis (exclude prehepatic causes)	Moderate: 25–50 μmol/L Severe: >50 μmol/L
Serum albumin level	Impaired hepatic protein synthesis	Moderate: 30–35g/L Severe: <30g/L
Prothrombin activity	Impaired hepatic protein synthesis	Moderate: 40–70% Severe: <40%

a The severity is classified according to Donelli et al., [20] with some modifications.

ULN = upper limit of normal.

However, regarding dose adaptation in patients with cancer, it has to be recognised that the administration guidelines discussed previously in this section focus on patients with liver cirrhosis or fibrosis and not on patients with increased transaminase levels and/or cholestasis, which are found frequently among patients treated with antineoplastic drugs. Since the majority of antineoplastic drugs are metabolised by the liver and are associated with severe dose-dependent toxicity, the question of whether the dose has to be adapted in a patient with increased transaminase levels and/or cholestasis is important (table I). The most prevalent liver disease in this group of patients is the presence of liver metastases, possibly resulting in cholestasis and/or portal hypertension. [10,18,19] Since many antineoplastic drugs are potentially hepatotoxic themselves, drug-induced liver disease may also be problematic in patients undergoing repetitive cycles of chemotherapy.

Therefore, the aims of the review were to: (i) categorise the antineoplastic drugs according to pharmacokinetic criteria; (ii) compare this categorisation with the dose recommendations in patients with liver disease given in the standard literature; (iii) formulate dose recommendations for dose adaptation; and (iv) localise gaps in the current recommendations.

## 2. Literature Search Methodology

We searched MEDLINE and EMBASE for studies dealing with dose adaptation and hepatic adverse effects for all antineoplastic drugs that were on the market in Switzerland at the end of 2004. The databases were screened using the following medical subject heading (MeSH) terms: 'antineoplastic agents', 'drug toxicity', 'pharmacokinetics' and 'liver diseases'. The references detected by this search were then hand screened for other suitable papers. In addition to databases, the standard literature was screened for dose-adaptation recommendations and adverse effects on the liver, including the *Swiss Compendium of Drugs*<sup>[22]</sup> (similar to the *Physicians' Desk Reference*<sup>[23]</sup>), *Therapeutic Drugs*<sup>[24]</sup> and *Hepatotoxicity*. [25]

The antineoplastic drugs were categorised according to pharmacokinetic principles as outlined in the previous section and based on the reviews of Huet and Villeneuve<sup>[16]</sup> and Delco et al.<sup>[1]</sup> The categorisation system used is based on the level of hepatic extraction, bioavailability or protein binding of the specific drugs (table II). Values for bioavailability and protein binding could be found either in the original articles<sup>[20,26-54]</sup> or in other sources.<sup>[22-24,55,56]</sup> For hepatic extraction, data in the literature are rare, making it necessary to estimate extraction from bioavailability (table II) or by the following equation (equation 4):

b Hepatocellular injury is defined according to Benichou.<sup>[21]</sup>

c Cholestasis is defined according to Benichou.[21]

$$E = \frac{Q_0 \times Cl_{sys}}{Q}$$

(Eq. 4)

where Q<sub>0</sub> is the extrarenal dose fraction (the fraction of a drug that is not excreted unchanged by the kidney), Cl<sub>sys</sub> is the systemic clearance of the drug (determined in plasma) and Q the plasma flow across the liver. The values for Q<sub>0</sub> and Cl<sub>sys</sub> were obtained from the literature<sup>[22-24,56]</sup> and Q was assumed to be 900 mL/min.

Dosage recommendations originate either from the original articles or from the manufacturer as published in the *Physicians' Desk Reference*<sup>[23]</sup> and/ or the *Swiss Compendium of Drugs*.<sup>[22]</sup>

Drug-induced liver disease was classified according to Benichou<sup>[21]</sup> and the severity of liver disease according to Donelli et al.<sup>[20]</sup> (see table I).

#### 3. Literature Search Results

We identified a total of 111 articles that were relevant to our review. [26-54,57-137] In 17 of these, drug metabolism data were reported, 49 contained hepatic adverse effects of antineoplastic agents and 45 were kinetic studies in patients and/or dose-adjustment recommendations.

Data on the 69 antineoplastic drugs on the Swiss market at the end of 2004 have been compiled in to table format and are available from http://pages.uinbas.ch/klinpharm/. From these 69 drugs, 15 fell into category 1, nine fell into category 2 and 25 fell into category 3 (table III). Twenty drugs could not be classified (category 4), demonstrating a lack of data about hepatic extraction and/or bioavailability.

Fifty-two of the 69 drugs have an extrarenal dose fraction  $(Q_0)$  value of >0.4 indicating that most

antineoplastic drugs are heavily metabolised and/or excreted by the bile. Seven drugs have a Q<sub>0</sub> value of  $\leq 0.4$  and for ten drugs, the Q<sub>0</sub> value could not be identified. For 25 drugs, metabolism by the CYP system is important, and 18 drugs are excreted to a significant extent (>5%) by the bile (vinca alkaloids, doxorubicin and derivatives, amsacrine, biculatamide, dactinomycin, estramustine, exemestan, irinotecan, imitanib, mitoxantrone, paclitaxel and topotecan). For 16 of these drugs, dose-adaptation recommendations are given according to the serum bilirubin level and/or activity of alkaline phosphatase (see table IV). For docetaxel, [20,22] doxorubicin, [112,122-127] idarubicin, [99,100] irinotecan, [102] mitoxantrone, [35-37] paclitaxel, [40-43] topotecan, [56,138] vincristine<sup>[139,140]</sup> and vinorelbine,<sup>[141]</sup> these recommendations are based on clinical studies. For amsacrine, bicalutamide, dactinomycin, daunorubicin, imatinib, vinblastine and vindesine, there are general statements in the Swiss Compendium of Drugs<sup>[22]</sup> and/or the Physicians' Desk Reference[23] that the dose should be adapted or stopped in patients with decreased liver function. For estramustine and exemestane, there are no recommendations regarding dosage in patients with liver disease. We recommend that these two drugs should be used with caution in patients with cholestasis and that the dose should be adapted according to dose-adapted adverse reactions.

For only 18 of the 69 drugs identified, recommendations for dose adaptation are based on published studies in patients with hepatic dysfunction. [27,28,31-34,40,41,47-52,54,110-120,122-137] For 44 of the 69 drugs, significant adverse effects on the liver have been reported. [26-28,31-34,40,41,47-54,110-137] This is important to realise, rendering drug-induced liver disease an important differential diagnosis in

Table II. Categorisation of antineoplastic drugs screened according to pharmacokinetic variables

Category	Level of hepatic extraction (%)	Resulting oral bioavailability
1	High (>60)	Oral bioavailability is <40% in the case of complete intestinal absorption (or accordingly lower, if intestinal absorption is not complete)
2	Intermediate (30-60)	Oral bioavailability is 40-70% in the case of complete intestinal absorption (or accordingly lower, if intestinal absorption is not complete)
3	Low (<30)	Oral bioavailability is >70% in the case of complete intestinal absorption (or accordingly lower, if intestinal absorption is not complete). In this category, protein binding may be relevant: for drugs with high binding to albumin (>90%), hepatic clearance may increase
4	Unknown	Not known

Table III. Antineoplastic drugs listed by hepatic extraction category

Category	Drugs
1	Capecitabine, cytarabine, docetaxel, doxorubicin, epirubicin, exemestane, fluorouracil, formestane, gemcitabine, idarubicin, medroxyprogesterone, mercaptopurine, mitoxantrone, vinblastine, vinorelbine
2	Bicalutamide, busulfan, cladribine, estramustine, gefitinib, irinotecan, melphalan, paclitaxel, topotecan
3	Aminoglutethimide, anastrozole, bleomycin, carboplatin, chlorambucil, cisplatin, cyclophosphamide, cyproterone, dacarbazine, etoposide, fludarabine, fosfestrol, goserelin, hydroxycarbamide (hydroxyurea), ifosfamide, imatinib, letrozole, leuprorelin, lomustine, methotrexate, temozolomide, thiotepa, toremifene, triptorelin, vincristine
4	Aldesleukin, alemtuzumab, amsacrine, buserelin, cetuximab, chlormethine (mechlorethamine), dactinomycin, daunorubicin, flutamide, megestrol, mitomycin, nimustine, oxaliplatin, raltitrexed, rituximab, tamoxifen, thioguanine, trastuzumab, tretinoin, vindesine

patients with malignant tumours and impaired hepatic function.

#### 4. Discussion

Our review demonstrates that for antineoplastic drugs, there is a discrepancy between the general recommendations of how drugs should be administered to patients with liver disease<sup>[1]</sup> and the available kinetic data for these drugs. The most important gaps are a lack of information regarding hepatic extraction and of kinetic studies for critical drugs in patients with impaired liver function.

As explained in the introduction, data about hepatic extraction are important for the classification of a specific drug regarding hepatic elimination in patients with chronic liver disease and liver cirrhosis in particular. It is evident that these data are difficult to obtain, especially the determination of hepatic extraction of a drug, necessitating an invasive procedure that is usually not performed before a drug is marketed. Bioavailability is only a surrogate for hepatic extraction, since a low bioavailability can originate from both a high hepatic extraction and/or a low intestinal absorption. For drugs with a low bioavailability (<40%), hepatic extraction should therefore be known, since this parameter is critical for rational drug administration in patients with impaired liver function, as previously discussed. To circumvent this invasive procedure in humans, a possibility would be to obtain data using perfused livers from animals, e.g. pigs. To the best of our knowledge, no data have been published so far comparing hepatic extraction data for critical drugs between animals (such as pigs) and humans. Another possibility is to estimate hepatic extraction using Q<sub>0</sub>, systemic drug clearance and hepatic plasma flow (equation 4). The values obtained with this technique are in satisfactory agreement with the bioavailability for most drugs, with some exceptions (see full table on http://pages.unibas.ch/klinpharm/).

Regarding antineoplastic agents, many of these drugs are used intravenously only, which partially explains the lack of data about oral bioavailability. Nonetheless, taking into account the high prevalence of patients with impaired hepatic function among those treated with these types of drugs, [138] these data should be available for all substances on the market.

Kinetic studies have been conducted in two conditions in particular, namely in patients with cholestasis (as suggested by an increased serum bilirubin level) and in patients with hepatic metastases. Regarding cholestasis, studies exist for most antineoplastic drugs with significant biliary elimination (see table IV). These studies resulted in quantitative recommendations for dose adaptation, for example in jaundiced patients according to their serum bilirubin level. However, to the best of our knowledge, these recommendations have not been validated by kinetic and dynamic studies (including the incidence and severity of dose-dependent adverse effects) in such patients. With cholestasis, it remains unclear whether the serum bilirubin level is the best parameter for predicting dose adaptation or whether the serum bile acid level and/or activity of alkaline phosphatase would be more suitable.

Regarding hepatic metastases, only few studies exist and they have generally not resulted in clear dose-adaptation recommendations. Since hepatic metastases can be associated with portal hypertension and possibly porto-caval shunts, [10,18] the situation can be similar to patients with liver cirrhosis. Oral administration of drugs with a high hepatic extraction should therefore be performed cautiously

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Table IV. Kinetic data, hepatic adverse effects and dose adjustment recommendations for the use of antineoplastic drugs with a significant biliary excretion (BE) in patients with liver disease

Drug	Cata	Kinetic parameters <sup>b,c,d</sup>	Frequency of hepatic adverse effects <sup>e</sup>	Dose-dependent adverse reactions	Studies performed and dose recommendations
Amsacrine	4	Metabolism: glutathion conjugation, BE >50% <sup>[22]</sup> Q <sub>0</sub> : 1 V <sub>d</sub> : 1.40 L/kg t <sub>½</sub> : 5h PB: 97%	Sporadic: cholestasis, hyperbilirubinaemia <sup>[25]</sup>	Myelosuppression, cardiotoxicity (arrhythmia), hypotonia, nausea and vomiting, alopecia, mucositis <sup>[22,24]</sup>	Recommendation: 50% dose reduction if serum bilirubin level >34 $\mu$ mol/L.[ <sup>26]</sup> Dose reduction (70% of normal dose) in patients with severe liver disease[ <sup>22,24]</sup>
Bicalutamide	2	Metabolism: oxidation (CYP), glucuronidation, BE 40% <sup>[22]</sup> Qo: ≈1 ty₂: 139h PB: 98% Cl <sub>sys</sub> : 500 mL/min E: 0.56	One case of fulminant liver failure <sup>[68]</sup>	Blocked androgenic action (hot flushes, breast tenderness, gynaecomastia, reduced libido and erectile function), diarrhoea <sup>[22-24]</sup>	Recommendations: stop treatment if transaminase levels >3 $\times$ ULN or in patients with hyperbilirubinaemia <sup>[22]</sup>
Dactinomycin	ı 4	Metabolism: BE 50–90% <sup>[24]</sup> Qo: 0.7 Vd: 12 L/kg t½: 36h	Rare: hepatocellular injury, steatosis, VOD <sup>[25]</sup>	Myelosuppression, nausea and vomiting, diarrhoea, mucositis, alopecia <sup>[23,24]</sup>	Recommendation: 50% dose reduction in patients with hyperbilirubinaemia. Increase gradually while monitoring dose-dependent toxicity <sup>[26]</sup>
Daunorubicin	4	Metabolism: reduction, BE 40% <sup>[24]</sup> Q <sub>0</sub> : 0.9 V <sub>d</sub> : 40 L/kg t <sub>/s</sub> : 27h	Rare: VOD when combined with radiation <sup>[25]</sup>	Myelosuppression, nausea and vomiting, mucositis, alopecia, cardiotoxicity, diarrhoea <sup>[23,24]</sup>	Recommendation: 25% dose reduction if serum bilirubin level 20–50 $\mu$ mol/L, 50% dose reduction if serum bilirubin level >50 $\mu$ mol/L [22,24]
Docetaxel	1	Metabolism: oxidation (CYP3A4). <sup>[24]</sup> BE 75%, 10% as intact drug <sup>[22,24]</sup> Qo: 1 Vd: 1.6 L/kg ty <sub>2</sub> : 11h PB: 95% Cl <sub>sys</sub> : 650 mL/min E: 0.72		Myelosuppression, nausea and vomiting, diarrhoea, sensory neuropathy, mucositis, alopecia, fluid retention syndrome <sup>[23,24]</sup>	Studies: population kinetic studies show a 25% reduction of clearance in patients with transaminase levels >1.5 $\times$ ULN and alkaline phosphatase level >2.5 $\times$ ULN. In patients with moderate liver injury/cholestasis, clearance was reduced by $27\%^{[22,24]}$ Recommendation: 25% dose reduction if transaminase levels >1.5 $\times$ ULN or alkaline phosphatase level >2.5 $\times$ ULN. Docetaxel should not be administered if serum bilirubin level is increased or transaminase levels >3.5 $\times$ ULN or alkaline phosphatase level >6 $\times$ ULN[22,24]
					ULN or alkaline phosphatase level >6 $\times$

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Orug	Cata	Kinetic parameters <sup>b,c,d</sup>	Frequency of hepatic adverse effectse	Dose-dependent adverse reactions	Studies performed and dose recommendations
Doxorubicin	1	Metabolism: reduction to doxorubicinol, sulfation, glucuronidation, BE 50% <sup>[20,24]</sup> Q <sub>0</sub> : 0.95 V <sub>d</sub> : 17 L/kg t <sub>/2</sub> : 26h PB: 80% F: 5% Cl <sub>sys</sub> : 1150 mL/min E: ≈1	Rare: in combination with etoposide, cyclophosphamide and cisplatin cholestasis and VOD <sup>[25]</sup>	Myelosuppression, nausea and vomiting, mucositis, alopecia, cardiotoxicity <sup>[23,24]</sup>	Studies: in five patients with disseminated sarcoma, myelotoxicity and doxorubicin serum levels correlated with hyperbilirubinaemia. [114] In patients with hepatocellular carcinoma, myelotoxicity and serum doxorubicin/ doxorubicinol levels correlated with hyperbilirubinaemia. [115,116] In 17 patients with hyperbilirubinaemia. [115,116] In 17 patients with liver metastases and moderate liver disease, kinetics of doxorubicin were not changed but the half-life of doxorubicinol increased. [117] In four patients with moderate liver disease, the half-life of doxorubicin was doubled. [118] In patients with liver metastases and a mild increase in transaminase or alkaline phosphatase levels, the kinetics and toxicity of doxorubicin were not changed [115,116,119,120] Recommendation: 50% dose reduction if serum bilirubin level 20–50 µmol/L. [22,24,26,121] Donelli et al. [20] advise dose reduction only if serum bilirubin level is >50 µmol/L
darubicin	1	Metabolism: oxidation, BE 8-17%[64.66] Qc:≈1 t/;: 15.2h PB: 96% F: 28% Cl <sub>sys</sub> : 2000 mL/min E: ≈1	Frequent: hepatocellular injury, hyperbilirubinaemia [22]	Myelosuppression, mucositis, alopecia, nausea and vomiting, diarrhoea, elevated liver enzyme levels, cardiotoxicity <sup>[23,24]</sup>	Studies: kinetics of idarubicin are not changed in patients with metastases $^{[133,134]}$ Recommendation: $50\%$ dose reduction if serum bilirubin level 20–34 $\mu mol/L$ . Contraindicated if serum bilirubin level >34 $\mu mol/L^{[22]}$
matinib	3	Metabolism: N-demethylation (CYP3A), BE 20% <sup>[22]</sup> Q <sub>0</sub> : 0.95 V <sub>d</sub> : 4.9 L/kg t <sub>/2</sub> : 18h PB: 95% E: 98%	Sporadic: hyperbilirubinaemia, hepatocellular injury <sup>[22]</sup>	Myelosuppression, oedema, myalgia, fatigue <sup>[22]</sup>	Recommendations: stop treatment if serum bilirubin level >3 $\times$ ULN or transaminase levels >5 $\times$ ULN <sup>[22]</sup>

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rug	Cata	Kinetic parameters <sup>b,c,d</sup>	Frequency of hepatic adverse effects <sup>e</sup>	Dose-dependent adverse reactions	Studies performed and dose recommendations
inotecan	2	Metabolism: esterases, glucuronidation, CYP3A4, BE 25% <sup>[22,65]</sup> Qo: 0.75 Vd: 75 L/kg t <sub>1/2</sub> : 10h PB: 65% Cl <sub>sys</sub> : 430 mL/min E: 0.36		Myelosuppression, cholinergic syndrome (diarrhoea), alopecia, nausea and vomiting, mucositis, pulmonary toxicity <sup>[22,23]</sup>	Study: in patients with gastrointestinal cancer and cholestasis, the AUC for SN-38 (active metabolite) was 50% increased (serum bilirubin level 1.1–1.5 × ULN) or 100% increased (>1.5 ULN) <sup>[135]</sup> Recommendation: if serum bilirubin level >1.5 × ULN or transaminase levels >5 × ULN, dose reduction according to dose-dependent toxicity. Contraindicated if serum bilirubin level >5 × ULN. <sup>[22]</sup> Dose of irinotecan should be 350 mg/m² in patients with serum bilirubin level 1.1–1.5 × ULN and 200 mg/m² when serum bilirubin level >1.5 × ULN <sup>[135]</sup>
ditoxantrone	1	Metabolism: mono- or dicarboxylation (inactive), BE $25\%^{[22]}$ Qo: $0.95$ Vd: $10-15$ L/kg ty/: $57h$ PB: $76\%$ Cl <sub>sys</sub> : $750$ mL/min E: $0.79$	Frequent: hepatocellular injury <sup>[25]</sup>	Myelosuppression, mucositis, nausea and vomiting, diarrhoea, menstrual disorders, neurological disorders, cardiotoxicity, alopecia, hepatotoxicity, nephrotoxicity <sup>[22,24]</sup>	Studies: clearance reduced by 50% in patients with moderate liver disease $^{[137]}$ Patients with serum bilirubin level <60 $\mu$ mol/L tolerate 14 mg/m², patients with serum bilirubin level >60 $\mu$ mol/L and bad performance status have higher mortality with this dosage $^{[27]}$ In patients with liver metastases, a half-life of mitoxantrone correlated with serum bilirubin level and cholestasis $^{[28]}$ Recommendation: dose adjustment (8 mg/m²) or contraindicated (bad performance status) in patients with serum bilirubin level >60 $\mu$ mol/L $^{[27]}$
Paclitaxel	2	Metabolism: CYP3A, CYP2C8, BE >5% <sup>[30]</sup> Qo: 0.95 Vd: 2.0 L/kg t½: 3h PB: 95% Cl <sub>sys</sub> : 380 mL/min E: 0.41	Sporadic: hepatocellular injury, cholestasis Rare: hyperbilirubinaemia, liver failure <sup>[22]</sup>	Myelosuppression, peripheral neuropathy, arthralgia, myalgia, hypotension, nausea and vomiting, diarrhoea, mucositis, cardiotoxicity (arrhythmias, bradycardia) <sup>[22,24]</sup>	Studies: liver disease/liver cirrhosis appears to be a risk factor for systemic toxicity. $^{[31,32]}$ Increased risk for myelosuppression in patients with increased transaminase levels and/or serum bilirubin level >25 $\mu$ mol/L. $^{[33]}$ In patients with increased transaminase levels (3–10 $\times$ ULN) and hyperbilirubinaemia (1.3–2 $\times$ ULN), clearance was decreased by $\approx\!40\%^{[34]}$ Recommendation: monitor patients with liver disease well for adverse effects. Do not administer in patients with decompensated liver disease $^{[22,34]}$

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Drug	Cata	Kinetic parameters <sup>b,c,d</sup>	Frequency of hepatic adverse effects <sup>e</sup>	Dose-dependent adverse reactions	Studies performed and dose recommendations
Topotecan	2	Metabolism: esterases, BE 20% <sup>[46]</sup> Q <sub>0</sub> : 0.6 V <sub>d</sub> : 1.9 L/kg t <sub>/s</sub> : 2.4h PB: 35% F: 32% Cl <sub>sys</sub> : 825 mL/min E: 0.55		Myelosuppression, nausea and vomiting, alopecia, mucositis, diarrhoea <sup>[22,23]</sup>	Studies: 14 patients with increased transaminase levels and/or hyperbilirubinaemia (some with cirrhosis) were treated with 1.5 mg/m². Topotecan clearance correlated with ICG clearance but no more adverse effects were observed in patients with liver disease. [47] On the other hand, two-thirds of patients with hepatocellular carcinoma treated with topotecan developed grade IV neutropenia [48] Recommendation: no dose adjustment for patients with hepatic dysfunction but monitor patients well for systemic toxicity [47]
Vinblastine	1	Metabolism: CYP3A4, BE >50%[ <sup>24]</sup> Q <sub>0</sub> : 1 Vd: 20 L/kg t/ <sub>2</sub> : 25h PB: 75% Cl <sub>sys</sub> : 865 mL/min E: 0.96		Myelosuppression, nausea and vomiting, alopecia, mucositis, neurotoxicity (peripheral and autonomic), inappropriate ADH secretion (SIADH)[22,24]	Recommendation: 50% dose reduction if serum bilirubin level >50 $\mu mol/L^{[22]}$
Vincristine	3	Metabolism: CYP3A4, BE 70% <sup>[24]</sup> Qo: 0.9 Vd: 8.0 L/kg ty <sub>6</sub> : 23h PB: 75% Cl <sub>sys</sub> : 140 mL/min E: 0.14		Myelosuppression, nausea and vomiting, alopecia, mucositis, neurotoxicity (peripheral and autonomic), inappropriate ADH secretion (SIADH)[22,24]	Studies: in the presence of cholestasis/ hyperbilirubinaemia half-life was prolonged. <sup>[51]</sup> In patients with leukaemia or lymphoma and cholestasis, AUC and toxicity were increased <sup>[52]</sup> Recommendation: 50% dose reduction if serum bilirubin >50 µmol/L. <sup>[22]</sup> Some authors advise 50% dose reduction also if alkaline phosphatase level is increased <sup>[26]</sup>
Vindesine	4	Metabolism: CYP3A, BE >20% <sup>[22,23]</sup> V <sub>d</sub> : 8.8 L/kg t <sub>1/2</sub> : 24h Cl <sub>sys</sub> : 17.5 mL/min		Myelosuppression, nausea and vomiting, alopecia, mucositis, neurotoxicity (peripheral and autonomic), inappropriate ADH secretion (SIADH)[22,24]	Recommendation: monitor patients for dose-dependent adverse effects. Dose may need to be adjusted in patients with hyperbilirubinaemia (see vincristine)[22]

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ole IV. Contd				
ug Cai	Cat <sup>a</sup> Kinetic parameters <sup>b,c,d</sup>	Frequency of hepatic adverse Dose-dependent adverse effects <sup>®</sup>	Dose-dependent adverse reactions	Studies performed and dose recommendations
orelbine 1	Metabolism: CYP 3A, BE 50%[24,53] Qo: 0.85 Vd: 75 L/kg t/s: 30h PB: 15% F: ≈40%		Myelosuppression, neurotoxicity (peripheral and autonomic), mucositis, alopecia, pulmonary toxicity <sup>[22,23]</sup>	Myelosuppression, neurotoxicity Studies: in 19 patients with liver metastases, peripheral and autonomic), clearance was reduced by 50% in patients with nucositis, alopecia, pulmonary >75% of the liver replaced by tumour. <sup>[54]</sup> Recommendation: 50% dose reduction if >75% of liver replaced by tumour <sup>[54]</sup> or if serum bilirubin level >34 μmol/L <sup>[53]</sup>

F = 40-70%as follows: category 1: high hepatic extraction [E >60%, F <40%]; category 2: intermediate hepatic extraction (E = 30-60%, category 3: low hepatic extraction (E <30%, F >70%); category 4: hepatic extraction not known. Drugs were categorised

The fraction metabolised or excreted by bile (1-Q<sub>0</sub>: fraction excreted unchanged by the kidney)

: For calculation, bodyweight was assumed to be 70kg.

Calculated as described in equation 4 of the main article.

e Frequent: >10% of patients treated; sporadic: 1-10%; rare: <1%.

ADH = antidiuretic hormone; AUC = area under the concentration-time curve; Cat = drug category; Clsys = systemic clearance; CYP = cytochrome P450; E = hepatic extraction; **Q**<sub>0</sub> = extrarenal dose fraction; **SIADH** = syndrome of = veno-occlusive disease distribution; EGFR = epidermal growth factor receptor; F = bioavailability; ICG = indocyanine green; PB = fraction bound to proteins; limit of normal; V<sub>d</sub> = volume of nappropriate antidiuretic hormone secretion; ty2 = dominant half-life;

and kinetic data for such drugs should be available in patients when such drugs are approved.

Treatment with antineoplastic agents can either lead to liver disease or, for drugs metabolised by the liver and/or excreted by the bile, to increased systemic toxicity in patients with liver disease. For such drugs, there is an additional type of toxicity that may be relevant. In several patients with chronic hepatitis B, the immunosuppressive effect of antineoplastic agents was associated with a flare up of their hepatitis due to increased replication of the hepatitis B virus. [139-145] Since this condition is potentially fatal, [141] but can be prevented by previous treatment or prophylaxis with antiviral agents, the immune status regarding hepatitis B should be known before treatment with antineoplastic drugs.

#### 5. Conclusion

In conclusion, there are currently considerable gaps in the data needed for safe administration of antineoplastic drugs in patients with decreased hepatic function. Drug authorities should urge pharmaceutical companies to provide data before these drugs are approved. Considering kinetics, in particular data about oral bioavailability and/or hepatic extraction should be known. For drugs with a predominant hepatic metabolism and/or excretion, the kinetics in patients with liver metastases and/or cholestasis should be known before marketing authorisation is provided.

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#### References

- Delco F, Tchambaz L, Schlienger R, et al. Dose adjustment in patients with liver disease. Drug Saf 2005; 28 (6): 529-45
- Herbert MF. Guide to drug dosage in hepatic disease. In: Holford NHG, editor. Drug data handbook. 3rd ed. Auckland: Adis International, 1998: 179
- 3. Bass NM, Williams RL. Guide to drug dosage in hepatic disease. Clin Pharmacokinet 1988; 15 (6): 396-420
- Westphal JF, Brogard JM. Drug administration in chronic liver disease. Drug Saf 1997; 17 (1): 47-73
- Verbeeck RK, Horsmans Y. Effect of hepatic insufficiency on pharmacokinetics and drug dosing. Pharm World Sci 1998; 20 (5): 183-92

- Reichen J. Assessment of hepatic function with xenobiotics. Semin Liver Dis 1995; 15 (3): 189-201
- Chawla Y, Santa N, Dhiman RK, et al. Portal hemodynamics by duplex Doppler sonography in different grades of cirrhosis. Dig Dis Sci 1998; 43 (2): 354-7
- Iwao T, Toyonaga A, Oho K, et al. Value of Doppler ultrasound parameters of portal vein and hepatic artery in the diagnosis of cirrhosis and portal hypertension. Am J Gastroenterol 1997; 92 (6): 1012-7
- Pare P, Talbot J, Hoefs JC. Serum-ascites albumin concentration gradient: a physiologic approach to the differential diagnosis of ascites. Gastroenterology 1983; 85 (2): 240-4
- Albillos A, Cuervas-Mons V, Millan I, et al. Ascitic fluid polymorphonuclear cell count and serum to ascites albumin gradient in the diagnosis of bacterial peritonitis. Gastroenterology 1990; 98 (1): 134-40
- George J, Liddle C, Murray M, et al. Pre-translational regulation of cytochrome P450 genes is responsible for disease-specific changes of individual P450 enzymes among patients with cirrhosis. Biochem Pharmacol 1995; 49 (7): 873-81
- George J, Murray M, Byth K, et al. Differential alterations of cytochrome P450 proteins in livers from patients with severe chronic liver disease. Hepatology 1995; 21 (1): 120-8
- Marcellin P, de Bony F, Garret C, et al. Influence of cirrhosis on lamotrigine pharmacokinetics. Br J Clin Pharmacol 2001; 51 (5): 410-4
- Macdonald JI, Wallace SM, Mahachai V, et al. Both phenolic and acyl glucuronidation pathways of diflunisal are impaired in liver cirrhosis. Eur J Clin Pharmacol 1992; 42 (5): 471-4
- Sonne J, Andreasen PB, Loft S, et al. Glucuronidation of oxazepam is not spared in patients with hepatic encephalopathy. Hepatology 1990; 11 (6): 951-6
- Huet PM, Villeneuve JP. Determinants of drug disposition in patients with cirrhosis. Hepatology 1983; 3 (6): 913-8
- 17. Shand DG. Hepatic circulation and drug disposition in cirrhosis. Gastroenterology 1979; 77 (1): 185-6
- Theodor E. Portal hypertension complicating liver involvement in metastatic carcinoma: a case report. Isr J Med Sci 1979; 15 (3): 285-7
- Huang JF, Little JM. Malignant jaundice. Aust N Z J Surg 1987;
   57 (12): 905-9
- Donelli MG, Zucchetti M, Munzone E, et al. Pharmacokinetics of anticancer agents in patients with impaired liver function. Eur J Cancer 1998; 34 (1): 33-46
- Benichou C. Criteria of drug-induced liver disorders. Report of an international consensus meeting. J Hepatol 1990; 11 (2): 272-6
- Morant J. Arzneimittelkompendium der Schweiz. Basel: Documed AG, 2004
- Sifton DW. Physicians' desk reference. 58th ed. Montvale (NJ): Medical Economics Company, 2004
- Dollery C, Boobis A, Rawlins M, et al. Therapeutic Drugs, 2 ed. Edinburgh: Churchill Livingstone, 1999
- Zimmerman HJ. Hepatotoxicity. 2 ed. Philadelphia (PA): Lippincott Williams & Wilkins, 1999
- Koren G, Beatty K, Seto A, et al. The effects of impaired liver function on the elimination of antineoplastic agents. Ann Pharmacother 1992; 26 (3): 363-71
- Chlebowski RT, Bulcavage L, Henderson IC, et al. Mitoxantrone use in breast cancer patients with elevated bilirubin. Breast Cancer Res Treat 1989; 14 (3): 267-74
- 28. Smyth JF, Macpherson JS, Warrington PS, et al. The clinical pharmacology of mitozantrone. Cancer Chemother Pharmacol 1986; 17 (2): 149-52
- Pendyala L, Creaven PJ. In vitro cytotoxicity, protein binding, red blood cell partitioning, and biotransformation of oxaliplatin. Cancer Res 1993; 53 (24): 5970-6

- Rowinsky EK, Cazenave LA, Donehower RC. Taxol: a novel investigational antimicrotubule agent. J Natl Cancer Inst 1990; 82 (15): 1247-59
- Chao Y, Chan WK, Birkhofer MJ, et al. Phase II and pharmacokinetic study of paclitaxel therapy for unresectable hepatocellular carcinoma patients. Br J Cancer 1998; 78 (1): 34-9
- Payne JY, Holmes F, Cohen PR, et al. Paclitaxel: severe mucocutaneous toxicity in a patient with hyperbilirubinemia. South Med J 1996; 89 (5): 542-5
- Venook AP, Egorin MJ, Rosner GL, et al. Phase I and pharmacokinetic trial of paclitaxel in patients with hepatic dysfunction: Cancer and Leukemia Group B 9264. J Clin Oncol 1998; 16 (5): 1811-9
- Panday VR, Huizing MT, Willemse PH, et al. Hepatic metabolism of paclitaxel and its impact in patients with altered hepatic function. Semin Oncol 1997; 24 (4 Suppl. 11): S11-34-S11-38
- Clarke SJ, Zalcberg J, Olver I, et al. Open label, multi-centre phase II study of raltitrexed ('Tomudex') in patients with inoperable squamous-cell carcinoma of head and neck. Ann Oncol 2000; 11 (2): 239-41
- Raderer M, Fiebiger W, Wrba F, et al. Fatal liver failure after the administration of raltitrexed for cancer chemotherapy: a report of two cases. Cancer 2000; 89 (4): 890-2
- Maruyama S, Hirayama C, Abe J, et al. Chronic active hepatitis and liver cirrhosis in association with combined tamoxifen/ tegafur adjuvant therapy. Dig Dis Sci 1995; 40 (12): 2602-7
- Pinto HC, Baptista A, Camilo ME, et al. Tamoxifen-associated steatohepatitis-report of three cases. J Hepatol 1995; 23 (1): 95-7
- Pratt DS, Knox TA, Erban J. Tamoxifen-induced steatohepatitis. Ann Intern Med 1995; 123 (3): 236
- Floren LC, Hebert MF, Venook AP, et al. Tamoxifen in liver disease: potential exacerbation of hepatic dysfunction. Ann Oncol 1998; 9 (10): 1123-6
- Martinez Cerezo FJ, Tomas A, Donoso L, et al. Controlled trial of tamoxifen in patients with advanced hepatocellular carcinoma. J Hepatol 1994; 20 (6): 702-6
- van Maanen MJ, Huitema AD, Beijen JH. Influence of comedicated drugs on the biotransformation of thioTEPA to TEPA and thioTEPA-mercapturate. Anticancer Res 2000; 20 (3A): 1711-6
- Lazarus HM, Reed MD, Spitzer TR, et al. High-dose i.v. thiotepa and cryopreserved autologous bone marrow transplantation for therapy of refractory cancer. Cancer Treat Rep 1987; 71 (7-8): 689-95
- Lee JL, Gooley T, Bensinger W, et al. Veno-occlusive disease of the liver after busulfan, melphalan, and thiotepa conditioning therapy: incidence, risk factors, and outcome. Biol Blood Marrow Transplant 1999; 5 (5): 306-15
- Przepiorka D, Khouri I, Thall P, et al. Thiotepa, busulfan and cyclophosphamide as a preparative regimen for allogeneic transplantation for advanced chronic myelogenous leukemia. Bone Marrow Transplant 1999; 23 (10): 977-81
- Herben VM, Schoemaker E, Rosing H, et al. Urinary and fecal excretion of topotecan in patients with malignant solid tumours. Cancer Chemother Pharmacol 2002; 50 (1): 59-64
- O'Reilly S, Rowinsky E, Slichenmyer W, et al. Phase I and pharmacologic studies of topotecan in patients with impaired hepatic function. J Natl Cancer Inst 1996; 88 (12): 817-24
- Wall JG, Benedetti JK, O'Rourke MA, et al. Phase II trial to topotecan in hepatocellular carcinoma: a Southwest Oncology Group study. Invest New Drugs 1997; 15 (3): 257-60
- Anttila M, Laakso S, Nylanden P, et al. Pharmacokinetics of the novel antiestrogenic agent toremifene in subjects with altered liver and kidney function. Clin Pharmacol Ther 1995; 57 (6): 628-35

- Muller FO, Terblanche J, Schall R, et al. Pharmacokinetics of triptorelin after intravenous bolus administration in healthy males and in males with renal or hepatic insufficiency. Br J Clin Pharmacol 1997; 44 (4): 335-41
- 51. Van den Berg HW, Desai ZR, Wilson R, et al. The pharmacokinetics of vincristine in man: reduced drug clearance associated with raised serum alkaline phosphatase and dose-limited elimination. Cancer Chemother Pharmacol 1982; 8 (2): 215-9
- Desai ZR, Van den Berg HW, Bridges JM, et al. Can severe vincristine neurotoxicity be prevented? Cancer Chemother Pharmacol 1982; 8 (2): 211-4
- Leveque D, Jehl F. Clinical pharmacokinetics of vinorelbine. Clin Pharmacokinet 1996; 31 (3): 184-97
- Robieux I, Sorio R, Borsatti E, et al. Pharmacokinetics of vinorelbine in patients with liver metastases. Clin Pharmacol Ther 1996; 59 (1): 32-40
- Hardman JG, Limbird LE, Gilman AG. The pharmacological basis of therapeutics. 10th ed. New York: McGraw-Hill, 2001
- Taeschner W, Vozeh S. Pharmacokinetic drug data. In: Holford NHG, editor. Drug data handbook. 3rd ed. Auckland: Aids International, 1998: 48
- Reynolds NA, Wagstaff AJ. Cetuximab: in the treatment of metastatic colorectal cancer. Drugs 2004; 64 (1): 109-18; discussion 119-21
- Balis FM, Holcenberg JS, Bleyer WA. Clinical pharmacokinetics of commonly used anticancer drugs. Clin Pharmacokinet 1983; 8 (3): 202-32
- Boddy AV, Yule SM. Metabolism and pharmacokinetics of oxazaphosphorines. Clin Pharmacokinet 2000; 38 (4): 291-304
- Camaggi CM, Strocchi E, Comparsi R, et al. Biliary excretion and pharmacokinetics of 4'epidoxorubicin (epirubicin) in advanced cancer patients. Cancer Chemother Pharmacol 1986; 18 (1): 47-50
- Gunnarsson PO, Andersson SB, Johansson SA, et al. Pharmacokinetics of estramustine phosphate (Estracyt) in prostatic cancer patients. Eur J Clin Pharmacol 1984; 26 (1): 113-9
- Clemett D, Lamb HM. Exemestane: a review of its use in postmenopausal women with advanced breast cancer. Drugs 2000; 59(6): 1279-96
- 63. Katchen B, Buxbaum S. Disposition of a new, nonsteroid, antiandrogen, alpha,alpha,alpha,trifluoro-2-methyl-4'-nitro-m-propionotoluidide (Flutamide), in men following a single oral 200mg dose. J Clin Endocrinol Metab 1975; 41 (2): 373-9
- Gillies HC, Herriott D, Liang R, et al. Pharmacokinetics of idarubicin (4-demethoxydaunorubicin; IMI-30; NSC 256439) following intravenous and oral administration in patients with advanced cancer. Br J Clin Pharmacol 1987; 23 (3): 303-10
- Lokiec F, du Sorbier BM, Sanderink GJ. Irinotecan (CPT-11) metabolites in human bile and urine. Clin Cancer Res 1996; 2 (12): 1943-9
- Tamassia V, Pacciarini MA, Moro E, et al. Pharmacokinetic study of intravenous and oral idarubicin in cancer patients. Int J Clin Pharmacol Res 1987; 7 (5): 419-26
- Lee FY, Workman P, Roberts JT, et al. Clinical pharmacokinetics of oral CCNU (lomustine). Cancer Chemother Pharmacol 1985; 14 (2): 125-31
- Chodak GW. Bicalutamide-associated fulminant hepatic failure. Urology 1997; 50 (6): 1027
- Umezawa H, Ishizuka M, Maeda K, et al. Studies on bleomycin. Cancer 1967; 20 (5): 891-5
- Morris LE, Guthrie TH. Busulfan-induced hepatitis. Am J Gastroenterol 1988; 83 (6): 682-3
- Underwood JC, Shahani RT, Blackburn EK. Jaundice after treatment of leukemia with busulphan. BMJ 1971; 1 (5748): 556-7

Blum JL. The role of capecitabine, an oral, enzymatically activated fluoropyrimidine, in the treatment of metastatic breast cancer. Oncologist 2001; 6 (1): 56-64

- Robert F, Ezekiel MP, Spencer SA, et al. Phase I study of antiepidermal growth factor receptor antibody cetuximab in combination with radiation therapy in patients with advanced head and neck cancer. J Clin Oncol 2001; 19 (13): 3234-43
- Patel SP, Nast CC, Adler SG. Chlorambucil-induced acute hepatic failure in a patient with membranous nephropathy. Am J Kidney Dis 2000; 36 (2): 401-4
- Mok CC, Wong WM, Shek TW, et al. Cumulative hepatotoxicity induced by continuous low-dose cyclophosphamide therapy. Am J Gastroenterol 2000; 95 (3): 845-6
- Gustafsson LL, Eriksson LS, Dahl ML, et al. Cyclophosphamide-induced acute liver failure requiring transplantation in a patient with genetically deficient debrisoquine metabolism: a causal relationship? J Intern Med 1996; 240 (5): 311-4
- Goldberg JW, Lidsky MD. Cyclophosphamide-associated hepatotoxicity. South Med J 1985; 78 (2): 222-3
- Blake JC, Sawyer Am, Dooley JS et al. Severe hepatitis caused by cyproterone acetate. Gut 1990; 31: 556-7
- Pinganaud G, Chaslerie A, Bourdel Marchasson I, et al. Cyproterone-induced hepatotoxicity. Ann Pharmacother 1995; 29 (6): 634
- Pu YS, Liu CM, Kao JH, et al. Antiandrogen hepatotoxicity in patients with chronic viral hepatitis. Eur Urol 1999; 36 (4): 293-7
- Migliari R, Muscas G, Murru M, et al. Antiandrogens: a summary review of pharmacodynamic properties and tolerability in prostate cancer therapy. Arch Ital Urol Androl 1999; 71 (5): 293-302
- Rollins BJ. Hepatic veno-occlusive disease. Am J Med 1986; 81
   (2): 297-306
- Houghton AN, Shafi N, Rickles FR. Acute hepatic vein thrombosis occurring during therapy for Hodgkin's disease: a case report. Cancer 1979; 44 (6): 2324-9
- Roila F, Crino L, Carloni G, et al. Cyproterone acetate: hepatotoxicity and prostatic cancer treatment. Ann Oncol 1993; 4 (8): 701
- Tran A, Housset C, Boboc B, et al. Etoposide (VP 16-213) induced hepatitis. Report of three cases following standarddose treatments. J Hepatol 1991; 12 (1): 36-9
- Cuevas Campos MA, Pareja Llorens G, Garcia Romero E, et al. Toxic hepatitis caused by flutamide [in Spanish]. Gastroenterol Hepatol 1998; 21 (10): 499-500
- Dourakis SP, Alexopoulou AA, Hadziyannis SJ. Fulminant hepatitis after flutamide treatment. J Hepatol 1994; 20 (3): 350-3
- Moller S, Iversen P, Franzmann MB. Flutamide-induced liver failure. J Hepatol 1990: 10 (3): 346-9
- Okaneya T, Murata Y, Kinebuchi Y. Fatal hepatic failure following hepatitis caused by flutamide: a case report [in Japanese]. Nippon Hinyokika Gakkai Zasshi 1999; 90 (5): 590-3
- Pontiroli L, Sartori M, Pittau S, et al. Flutamide-induced acute hepatitis: investigation on the role of immunoallergic mechanisms. Ital J Gastroenterol Hepatol 1998; 30 (3): 310-4
- Wada T, Ueda M, Abe K, et al. Risk factor of liver disorders caused by flutamide--statistical analysis using multivariate logistic regression analysis [in Japanese]. Hinyokika Kiyo 1999; 45 (8): 521-6
- Satoh T, Egawa S, Katsuta M, et al. A case of fulminant hepatitis caused by antiandrogen, flutamide in a patient with prostate cancer. Nippon Hinyokika Gakkai Zasshi 1997; 88 (7): 694-6
- Wietzke P, Munke H, Hartmann H, et al. Hepatotoxicity of flutamide. Z Gastroenterol 1997; 35 (8): 631-5

- 94. Chapoutot C, Perney P, Le Bricquir Y, et al. Acute cytolytic hepatitis caused by hydroxycarbamide. Gastroenterol Clin Biol 1997; 21 (1): 87-9
- Gross R, Scapa E. Hepatotoxicity of 6-mercaptopurine in Crohn's disease. Am J Gastroenterol 1992; 87 (12): 1885-6
- Gross R. Hepatotoxicity of 6-mercaptopurine and azathioprine. Mayo Clin Proc 1994; 69 (5): 498
- Laidlaw ST, Reilly JT, Suvarna SK. Fatal hepatotoxicity associated with 6-mercaptopurine therapy. Postgrad Med J 1995; 71
  (840): 639
- Berkovitch M, Matsui D, Zipursky A, et al. Hepatotoxicity of 6-mercaptopurine in childhood acute lymphocytic leukemia: pharmacokinetic characteristics. Med Pediatr Oncol 1996; 26 (2): 85-9
- Gilbert SC, Klintmalm G, Menter A, et al. Methotrexate-induced cirrhosis requiring liver transplantation in three patients with psoriasis. A word of caution in light of the expanding use of this 'steroid-sparing' agent. Arch Intern Med 1990; 150 (4): 889-91
- Hakim NS, Kobienia B, Benedetti E, et al. Methotrexate-induced hepatic necrosis requiring liver transplantation in a patient with rheumatoid arthritis. Int Surg 1998; 83 (3): 224-5
- 101. Malatjalian DA, Ross JB, Williams CN, et al. Methotrexate hepatotoxicity in psoriatics: report of 104 patients from Nova Scotia, with analysis of risks from obesity, diabetes and alcohol consumption during long term follow-up. Can J Gastroenterol 1996; 10 (6): 369-75
- 102. Shergy WJ, Polisson RP, Caldwell DS, et al. Methotrexateassociated hepatotoxicity: retrospective analysis of 210 patients with rheumatoid arthritis. Am J Med 1988; 85 (6): 771-4
- ter Borg EJ, Seldenrijk CA, Timmer R. Liver cirrhosis due to methotrexate in a patient with rheumatoid arthritis. Neth J Med 1996; 49 (6): 244-6
- 104. West SG. Methotrexate hepatotoxicity. Rheum Dis Clin North Am 1997; 23 (4): 883-915
- Whiting-O'Keefe QE, Fye KH, Sack KD. Methotrexate and histologic hepatic abnormalities: a meta-analysis. Am J Med 1991; 90 (6): 711-6
- Soh LT, Ang PT, Sng I, et al. Fulminant hepatic failure in non-Hodgkin lymphoma patients treated with chemotherapy. Eur J Cancer 1992; 28A (8-9): 1338-9
- 107. Farrow AC, Buchanan GR, Zwiener RJ, et al. Serum aminotransferase elevation during and following treatment of childhood acute lymphoblastic leukemia. J Clin Oncol 1997; 15 (4): 1560-6
- 108. Exadaktylos P, Reiss T, Schobess R, et al. Acute hepatotoxicity with intermediate-dose methotrexate in children with leukemia and non-Hodgkin's lymphoma [in German]. Klin Padiatr 1994; 206 (4): 315-8
- Fabbri A, Motta E, Ferrari S, et al. High-dose methotrexate treatment and liver function in patients with osteosarcoma. J Intern Med 1994; 236 (2): 209-14
- 110. Wagner T, Heydrich D, Bartels H, et al. Effect of damaged liver parenchyma, renal insufficiency and hemodialysis on the pharmacokinetics of cyclophosphamide and its activated metabolites [in German]. Arzneimittelforschung 1980; 30 (9): 1588-92
- 111. Twelves C, Glynne-Jones R, Cassidy J, et al. Effect of hepatic dysfunction due to liver metastases on the pharmacokinetics of capecitabine and its metabolites. Clin Cancer Res 1999; 5 (7): 1696-702
- 112. Twelves CJ, O'Reilly SM, Coleman RE, et al. Weekly epirubicin for breast cancer with liver metastases and abnormal liver biochemistry. Br J Cancer 1989; 60(6): 938-41
- Juma FD. Effect of liver failure on the pharmacokinetics of cyclophosphamide. Eur J Clin Pharmacol 1984; 26 (5): 591-3

- Benjamin RS. Pharmacokinetics of adriamycin (NSC-123127) in patients with sarcomas. Cancer Chemother Rep 1974; 58
   (2): 271-3
- 115. Chan KK, Chlebowski RT, Tong M, et al. Clinical pharmacokinetics of adriamycin in hepatoma patients with cirrhosis. Cancer Res 1980; 40 (4): 1263-8
- 116. Johnson PJ, Dobbs N, Kalayci C, et al. Clinical efficacy and toxicity of standard dose adriamycin in hyperbilirubinaemic patients with hepatocellular carcinoma: relation to liver tests and pharmacokinetic parameters. Br J Cancer 1992; 65 (5): 751-5
- 117. Preiss R, Matthias M, Sohr R, et al. Pharmacokinetics of adriamycin, adriamycinol, and antipyrine in patients with moderate tumor involvement of the liver. J Cancer Res Clin Oncol 1987; 113 (6): 593-8
- Piscitelli SC, Rodvold KA, Rushing DA, et al. Pharmacokinetics and pharmacodynamics of doxorubicin in patients with small cell lung cancer. Clin Pharmacol Ther 1993; 53 (5): 555-61
- Morris RG, Reece PA, Dale BM, et al. Alteration in doxorubicin and doxorubicinol plasma concentrations with repeated courses to patients. Ther Drug Monit 1989; 11 (4): 380-3
- Mross K, Maessen P, van der Vijgh WJ, et al. Pharmacokinetics and metabolism of epidoxorubicin and doxorubicin in humans. J Clin Oncol 1988; 6 (3): 517-26
- Dobbs NA, Twelves CJ. Anthracycline doses in patients with liver dysfunction: do UK oncologists follow current recommendations? Br J Cancer 1998; 77 (7): 1145-8
- 122. Camaggi CM, Strocchi E, Tamassia V, et al. Pharmacokinetic studies of 4'-epi-doxorubicin in cancer patients with normal and impaired renal function and with hepatic metastases. Cancer Treat Rep 1982; 66 (10): 1819-24
- 123. Jakobsen P, Bastholt L, Dalmark M, et al. A randomized study of epirubicin at four different dose levels in advanced breast cancer. Feasibility of myelotoxicity prediction through single blood-sample measurement. Cancer Chemother Pharmacol 1991; 28 (6): 465-9
- 124. Speth PA, Linssen PC, Beex LV, et al. Cellular and plasma pharmacokinetics of weekly 20-mg 4'-epi-adriamycin bolus injection in patients with advanced breast carcinoma. Cancer Chemother Pharmacol 1986; 18 (1): 78-82
- Dobbs NA, Twelves CJ, Rizzi P, et al. Epirubicin in hepatocellular carcinoma: pharmacokinetics and clinical activity. Cancer Chemother Pharmacol 1994; 34 (5): 405-10
- 126. Twelves CJ, Richards MA, Smith P, et al. Epirubicin in breast cancer patients with liver metastases and abnormal liver biochemistry: initial weekly treatment followed by rescheduling and intensification. Ann Oncol 1991; 2 (9): 663-6
- 127. Twelves CJ, Dobbs NA, Michael Y, et al. Clinical pharmacokinetics of epirubicin: the importance of liver biochemistry tests. Br J Cancer 1992; 66 (4): 765-9
- D'Incalci M, Rossi C, Zucchetti M, et al. Pharmacokinetics of etoposide in patients with abnormal renal and hepatic function. Cancer Res 1986; 46 (5): 2566-71
- Hande KR, Wolff SN, Greco FA, et al. Etoposide kinetics in patients with obstructive jaundice. J Clin Oncol 1990; 8 (6): 1101-7
- Joel SP, Shah R, Clark PI, et al. Predicting etoposide toxicity: relationship to organ function and protein binding. J Clin Oncol 1996; 14 (1): 257-67
- Aita P, Robieux I, Sorio R, et al. Pharmacokinetics of oral etoposide in patients with hepatocellular carcinoma. Cancer Chemother Pharmacol 1999; 43 (4): 287-94
- Fleming RA, Milano GA, Etienne MC, et al. No effect of dose, hepatic function, or nutritional status on 5-FU clearance following continuous (5-day), 5-FU infusion. Br J Cancer 1992; 66 (4): 668-72

 Lu K, Savaraj N, Kavanagh J, et al. Clinical pharmacology of 4-demethoxydaunorubicin (DMDR). Cancer Chemother Pharmacol 1986; 17 (2): 143-8

- 134. Camaggi CM, Strocchi E, Carisi P, et al. Idarubicin metabolism and pharmacokinetics after intravenous and oral administration in cancer patients: a crossover study. Cancer Chemother Pharmacol 1992; 30 (4): 307-16
- Raymond E, Boige V, Faivre S, et al. Dosage adjustment and pharmacokinetic profile of irinotecan in cancer patients with hepatic dysfunction. J Clin Oncol 2002; 20 (21): 4303-12
- 136. Skoglund KA, Soderhall S, Beck O, et al. Plasma and urine levels of methotrexate and 7-hydroxymethotrexate in children with ALL during maintenance therapy with weekly oral methotrexate. Med Pediatr Oncol 1994; 22 (3): 187-93
- Savaraj N, Lu K, Manuel V, et al. Pharmacology of mitoxantrone in cancer patients. Cancer Chemother Pharmacol 1982; 8

   (1): 113-7
- O'Reilly SM, Richards MA, Rubens RD. Liver metastases from breast cancer: the relationship between clinical, biochemical and pathological features and survival. Eur J Cancer 1990; 26 (5): 574-7
- 139. Sato T, Kato J, Kawanishi J, et al. Acute exacerbation of hepatitis due to reactivation of hepatitis B virus with mutations in the core region after chemotherapy for malignant lymphoma. J Gastroenterol 1997; 32 (5): 668-71
- 140. Yoshiba M, Sekiyama K, Sugata F, et al. Reactivation of precore mutant hepatitis B virus leading to fulminant hepatic

- failure following cytotoxic treatment. Dig Dis Sci 1992; 37 (8): 1253-9
- Yeo W, Steinberg JL, Tam JS, et al. Lamivudine in the treatment of hepatitis B virus reactivation during cytotoxic chemotherapy. J Med Virol 1999; 59 (3): 263-9
- Dai MS, Lu JJ, Chen YC, et al. Reactivation of precore mutant hepatitis B virus in chemotherapy-treated patients. Cancer 2001; 92 (11): 2927-32
- 143. Faggioli P, De Paschale M, Tocci A, et al. Acute hepatic toxicity during cyclic chemotherapy in non Hodgkin's lymphoma. Haematologica 1997; 82 (1): 38-42
- 144. Lok AS, Liang RH, Chiu EK, et al. Reactivation of hepatitis B virus replication in patients receiving cytotoxic therapy. Report of a prospective study. Gastroenterology 1991; 100 (1): 182-8
- 145. Yoshiba M, Sekiyama K, Iwabuchi S, et al. Recurrent fulminant hepatic failure in an HB carrier after intensive chemotherapy. Dig Dis Sci 1993; 38 (9): 1751-5

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